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Rapid mapping of schistosomiasis and other neglected tropical diseases in the context of integrated control programmes in Africa

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Abstract

SUMMARY—There is growing interest and commitment to the control of schistosomiasis and other so-called neglected tropical diseases (NTDs). Resources for control are inevitably limited, necessitating assessment methods that can rapidly and accurately identify and map high-risk communities so that interventions can be targeted in a spatially-explicit and cost-effective manner. Here, we review progress made with (i) mapping schistosomiasis across Africa using available epidemiological data and more recently, climate-based risk prediction; (ii) the development and use of morbidity questionnaires for rapid identification of high-risk communities of urinary schistosomiasis; and (iii) innovative sampling-based approaches for intestinal schistosomiasis, using the lot quality assurance sampling technique. Experiences are also presented for the rapid mapping of other NTDs, including onchocerciasis, loiasis and lymphatic filariasis. Future directions for an integrated rapid mapping approach targeting multiple NTDs simultaneously are outlined, including potential challenges in developing an integrated survey tool. The lessons from the mapping of human helminth infections may also be relevant for the rapid mapping of malaria as its control efforts are intensified.

Keywords

schistosomiasis; neglected tropical diseases; geographical information system; rapid mapping; morbidity questionnaires; lot quality assurance sampling; integrated control; Africa

INTRODUCTION

The control of schistosomiasis (*Schistosoma haematobium* and *S. mansoni*) and soil-transmitted helminthiasis (STH: *Ascaris lumbricoides*, *Trichuris trichiura* and hookworm) has been greatly boosted since the World Health Assembly, in May 2001, resolved to regularly treat at least 75% of school-aged children and other high-risk groups by 2010 (WHO, 2002). In parallel, there has been an increase in political, financial and technical support for helminth control, with a focus on preventive chemotherapy using praziquantel against schistosomiasis and albendazole or mebendazole against STH (Utzinger *et al.* 2003; WHO, 2006). In particular,

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school-based programmes have been shown to be a cost-effective approach for controlling schistosomiasis and STH (Partnership for Child Development, 1997; Bundy *et al.* 2006; Brooker *et al.* 2008). The increase in school participation worldwide as one of the key millennium development goals (MDGs), as well as the specific promotion of school health programmes as part of these efforts, provides an additional strong policy incentive to deliver deworming through school systems. Identifying which schools and communities require treatment is an essential part of any control programme, and a key issue for epidemiological mapping.

For schistosomiasis surveys, recommendations put forth by the World Health Organization (WHO) are based on the random selection of at least 50 individuals within defined ecological zones using the 'gold' standard of parasitological diagnosis, namely light microscopy of repeated stool and/or urine examinations to detect and quantify distinctive schistosome eggs (Bergquist *et al.* 2009; Dansoh-Appiah *et al.* 2009). However, the costs of such an approach are high and health systems often lack the necessary equipment, supplies and skilled personnel (Kurowski *et al.* 2007). The technical capacity to conduct population-based surveys is also often limited (Mathieu *et al.* 2003), necessitating low-cost, simple and rapid alternatives. This operational requirement is also true for a number of other tropical diseases.

In the last five years, there has been an expansion of interest in a range of so-called neglected tropical diseases (NTDs), including lymphatic filariasis, onchocerciasis, trachoma as well as schistosomiasis and STH (Molyneux *et al.* 2005; Lammie *et al.* 2006; Utzinger and de Savigny, 2006; WHO, 2006; Hotez *et al.* 2007). Control of these diseases is mainly through mass drug administration (MDA) with annual or semi-annual doses of safe and efficacious oral treatments. Given the similarity of approaches to implementation, it is suggested that disease-specific programmes can enhance their efficiencies and cost-effectiveness by delivering drugs in a coordinated fashion (WHO, 2006; Hotez *et al.* 2007). WHO guidelines for preventive chemotherapy provide a useful framework for countries that are implementing integrated NTD control (WHO, 2006). An important first step is to define the geographical distribution and degree of overlap of different NTDs in order to define treatment strategies (Brady *et al.* 2006; Kolaczinski *et al.* 2007); simple, cheap and rapid methods of assessment are clearly necessary for initial mapping.

This review examines how a number of scientific approaches can be used to rapidly and effectively map the distribution of schistosomiasis in Africa. We also focus on the operational experience of mapping other NTDs, and on the potential integration of rapid mapping approaches in the implementation of combined NTD control. Finally, we suggest that experiences and expertise gained from the rapid mapping of schistosomiasis and other NTDs may be relevant for rapid assessment of malaria.

IDENTIFYING AREAS OF SCHISTOSOMIASIS RISK

Disease mapping and climate-based risk models

Over the years, thousands of surveys have been undertaken to estimate the prevalence of schistosomiasis in different locations across Africa (Doumenge *et al.* 1987; Brooker *et al.* 2000). While there are problems regarding comparability between surveys due to differences in sampling methods, age groups covered, diagnostic approaches taken, and timings of surveys, an 'Atlas of Human Helminth Infections' has attempted to extract and collate relevant surveys into a single database, with the aim of describing the distribution of infection across the African continent (Brooker *et al.* 2000). This work served to highlight the tremendous paucity of empirical data for large areas of the African continent, with survey data on schistosomiasis available in 2000 for only a third of districts in Africa. Since 2000, a large number of national surveys have been undertaken, and current work is ongoing to update the atlas as well as to

geo-position surveys to actual locations. Figure 1 presents an example of current progress for East and Southern Africa.

To help overcome the paucity of survey data, research efforts have been made to predict schistosome infection risk at non-sampled locations on the basis of climate-based risk models, using geographical information system (GIS), remote sensing (RS) and geostatistics (for recent reviews see: Brooker, 2007; Simoonga *et al.* 2009; Vounatsou *et al.* 2009). Although useful to guide control efforts at national scales by excluding areas where no or limited transmission occurs, such risk models are unable to confine the geographical variation of infection evident at community levels. This is because schistosomiasis is a highly focal disease, such that control needs to be targeted on a community by community basis. This requirement will be increasingly important as national control programmes reduce the overall level of infection, and morbidity and spatial patterns potentially become even more heterogeneous, which has important repercussions for control (Raso *et al.* 2007). Below we consider a number of approaches to define community patterns of schistosomiasis, and discuss their implementation in practice.

Morbidity questionnaires

The signs and symptoms of urinary schistosomiasis, such as blood in urine and pain when urinating, are clearly recognised by individuals, and it is not common for school-aged children to report having blood in urine without having urinary schistosomiasis. Such specificity of this sign of *S. haematobium* infection in endemic areas has allowed blood in urine to be incorporated into morbidity questionnaires which identify communities with a high prevalence of infection and warranting mass treatment (WHO, 1995). The use of reported blood in urine to identify high prevalence communities using a simple school-based questionnaire was first developed and administered through the education system in Kilombero and Kilosa districts of Tanzania (Lengeler *et al.* 1991a, b). A number of studies have subsequently demonstrated that morbidity questionnaires can rapidly and cost-effectively identify high-risk communities (for a review see: Lengeler *et al.* 2002). These studies confirm that morbidity questionnaires administered through the education system are accurate and allow for rapid and cost-effective identification of high-risk communities of urinary schistosomiasis (Ansell *et al.* 2002; Lengeler *et al.* 2002). At an operational level, two aspects are worth highlighting. First, the recall period length (2 weeks *versus* 4 weeks) had only a minor effect on the diagnostic performance of the questionnaire (van der Werf *et al.* 2003). Second, age and sex influence the reliability of self-reported schistosomiasis as a diagnostic method (Ansell *et al.* 2001; Clements *et al.* 2008a).

Questionnaire surveys for urinary schistosomiasis are now recommended as a first step in implementing national schistosomiasis control programmes. In the late 1990s, for example, questionnaires were administered to 1291 schools in five regions of Côte d'Ivoire. Within 5 weeks filled-in questionnaires were returned by 1091 schools (84.5%) with 118,327 schoolchildren being interviewed, with a recent history of blood in urine being reported by 22,035 (18.6%) of the children. Based on these results, 186 schools were considered at high risk of urinary schistosomiasis (N'Guessan *et al.* 2007). In Tanzania in 2004, a morbidity questionnaire was distributed through the existing infrastructure of the Ministry of Education, to all schools in the country and teachers were trained in how to complete the questionnaire. More than 2.5 million schoolchildren in 12,399 schools were interviewed and results on the frequencies of self-reported blood in urine and self-reported schistosomiasis were utilized to guide the spatial targeting of praziquantel administration (Clements *et al.* 2008b).

Considerable efforts have also been made to extend the questionnaire approach from urinary to intestinal schistosomiasis. These efforts were predicated on the results of numerous epidemiological and hospital-based studies showing that *S. mansoni* infections, especially those with a moderate or heavy intensity (>100 eggs per gram of stool), are associated with a recent history of blood in stool and/or (bloody) diarrhoea (Gryseels, 1992; Utzinger *et al.*

1998), and that these symptoms are valuable markers for identification of high-risk communities of intestinal schistosomiasis (Utzingler *et al.* 2000; Lengeler *et al.* 2002). However, compared to urinary schistosomiasis, the questionnaire approach for intestinal schistosomiasis only showed a moderate sensitivity. It was thus concluded that parasitological validation should precede any large-scale use of questionnaires for rapid screening of intestinal schistosomiasis in a given epidemiological setting (Brooker *et al.* 2001; Lengeler *et al.* 2002).

Rapid mapping of intestinal schistosomiasis

Concerns about the lack of a reliable questionnaire approach for intestinal schistosomiasis and the need for further validation prompted researchers to explore alternative ways to reduce the sampling effort required to assess the prevalence and distribution of *S. mansoni*. Lot quality assurance sampling (LQAS) is an approach that relies on a relatively limited number of observations to classify sampling units so that a decision can be rapidly reached. Originally developed by industry to monitor production quality, it has been increasingly adapted for use in population health surveys (Robertson and Valadez, 2006). Recent operational research has highlighted the potential of LQAS for identifying high-risk communities for *S. mansoni* (Rabarijaona *et al.* 2001; Brooker *et al.* 2005). In this approach, a specified number of individuals are randomly selected to provide stool samples which are examined until either a predetermined number of infected individuals is identified, i.e. indicating a high prevalence ($\geq 50\%$), or the specified number of individuals sampled without the threshold being reached, i.e. indicating a prevalence below 50% (WHO, 2002). A finer classification system may be provided by applying alternative, secondary sampling plans to data already collected and classified, by previously applied sampling plans, as coming from communities with a prevalence below 50%, and used to distinguish between moderate (20-49%) and low (<20%) prevalence schools (Brooker *et al.* 2005). The approach has the benefit of allowing the survey to stop as soon as it has sufficient information about the likely category of *S. mansoni* prevalence and facilitates an intervention strategy to be planned, e.g. mass treatment of entire communities with praziquantel where prevalence exceeds 50%.

In Uganda, a comprehensive investigation, employing a combination of computer simulations and field studies, showed that LQAS provides a highly reliable approach to classifying schools according to treatment strategy (Brooker *et al.* 2005). Cost-effectiveness analysis indicated that screening and treating schools on the basis of LQAS was more cost-effective than mass treating all schools without prior screening in a given district. On the basis of these results, the Ugandan national helminth control programme determined in 2006, to undertake nationwide rapid mapping survey using LQAS to assess the status of *S. mansoni* following several treatment rounds. The purpose of the investigation was to classify schools according to different prevalence thresholds: <20%, 20-50% and >50%, in order to direct the future treatment strategy for the national control programme. Comparing these rapid mapping results against parasitological data collected prior to treatment (Kabatereine *et al.* 2004), it can be seen that greatest apparent change occurred along the Nile basin in the north-western region of the country and along the shores of Lake Albert (Figure 2). The results also highlight that areas of high prevalence of *S. mansoni* still exist and reveals additional districts where treatment is warranted. Although illustrative, such mapping results cannot definitively assess programme impact – a robust analysis of programme impact is presented elsewhere (Kabatereine *et al.* 2007) – but can provide useful qualitative information for planning purposes as control programmes roll out. The overall estimated cost of the rapid mapping investigation was US\$ 21,182, illustrating the low cost of the approach to rapidly assess the distribution of intestinal schistosomiasis at a national level. The approach could be further simplified if combined with rapid, yet reliable diagnosis, which we now consider.

Rapid diagnosis of schistosomiasis

Reliable and rapid diagnosis is central to the mapping of schistosomiasis. The 'gold' standard for diagnosing schistosomiasis, particularly in the early stages of control when morbidity reduction is the declared objective, is the detection of distinctive eggs in urine by the filtration method (for *S. haematobium*) or in stool by the Kato-Katz technique or other direct methods (for *S. mansoni*) (Bergquist *et al.* 2009). However, stool examination suffers day-to-day and intra-specimen variation in egg output necessitating sample collection over consecutive days to ensure maximal sensitivity (de Vlas and Gryseels, 1992; Engels *et al.* 1996, 1997; Utzinger *et al.* 2001). Moreover, urine filtration and Kato-Katz thick smear examination require expert microscopy.

For *S. haematobium*, testing urine with reagent strips for blood is a simple, indirect method of identifying infected individuals and who require treatment (Mafe, 1997; Lengeler *et al.* 2002). A wealth of studies report high sensitivity and specificity of reagent strips when compared to urine filtration when both methods were performed by research teams (Table 1). Furthermore, studies show that training of teachers (usually within half a day) allow for subsequent reagent strip testing by the teachers, resulting in moderate to high sensitivity and specificity (Table 2).

One limitation of haematuria reagent strips is that they can be affected by factors such as menstruation and genitourinary infections (Hall and Fentiman, 1999; Poggensee *et al.* 2000). A further shortcoming is that they are semi-quantitative and have a limited concentration detection range. Such limitations have prompted researchers to investigate alternative rapid diagnosis methods. In recent years, the diagnosis of many tropical diseases, including malaria, has been greatly enhanced by the use of easy-to-use and accurate immunological tests, where relevant parasite antigens are detected in finger-prick blood (Moody, 2002; Marx *et al.* 2005). Although accurate, non-invasive rapid antigen detection assays are currently available for lymphatic filariasis, onchocerciasis and visceral leishmaniasis (Weil *et al.* 1997; Ayong *et al.* 2005; Chappuis *et al.* 2006), progress in this area for schistosomiasis has been slow.

Schistosome antigens are readily detectable in sera, but sample collection requiring venopuncture is often unpractical, not widely accepted, and requires trained personnel. An alternative diagnostic method is based on schistosome circulating cathodic antigens (CCA), which are excreted in both serum and urine. For practical convenience, a urine-based test has sought to be developed. The first such test was an enzyme-linked immunosorbent assay (ELISA), which led to a dipstick format (van Etten *et al.* 1997). More recent attention has focused upon immuno-lateral flow principles, which utilise capillary diffusion of reagents through nitrocellulose strips (van Dam *et al.* 2004). A CCA-based dipstick became commercially available in 2003 from BV European Veterinary Laboratory. Field evaluations in Burkina Faso, Niger, Uganda and Zanzibar showed that the test was insensitive in detecting urinary schistosomiasis but highly (80%) sensitive and specific in the detection of intestinal schistosomiasis (Stothard *et al.* 2006). Studies in Ethiopia confirmed the high sensitivity and specificity of CCA-based dipsticks for *S. mansoni* but showed a lower sensitivity in low transmission settings (Legesse and Erko, 2007, 2008). Other work in Ethiopia found a low sensitivity (52%) and specificity (64%) for the detection of *S. haematobium* (Ayele *et al.* 2008), thereby supporting the claim by Stothard and colleagues that CCA-based dipsticks have no or limited value for detection for *S. haematobium*. A current limitation of using CCA dipsticks for *S. mansoni* is the prohibitively expensive cost, which is presently at US\$ 2.6 per test.

RAPID MAPPING OF OTHER NTDS

The rapid assessment of schistosomiasis has been able to successfully build upon rapid mapping methods developed for other NTDS. The first such method was for onchocerciasis, a disease caused by infection with a parasitic filarial worm, *Onchocerca volvulus*. Control of onchocerciasis involves the yearly mass community-directed distribution of ivermectin as part of the African Programme for Onchocerciasis Control (APOC). Priority areas for mass distribution of ivermectin are identified on the basis of the rapid epidemiological mapping of onchocerciasis (REMO) method, which takes into consideration specific spatio-epidemiological characteristics of the disease (spatial distribution of vectors in breeding sites along fast-flowing and well oxygenated rivers) to identify areas likely to be at high risk (Ngoumou *et al.* 1994; Katarbarwa *et al.* 1999). A subsample of communities in identified areas are then screened using the rapid epidemiological assessment for onchocerciasis (REA) method, which is based on the estimation of the prevalence of onchocercal nodules in adult males using simple palpation. Results of REMO have been effectively incorporated into a GIS to visualize priority areas for mass distribution of ivermectin and estimate the number of individuals to be treated, and has greatly helped APOC to prioritize allocation of resources according to need (Noma *et al.* 2002).

The approach has also been extended to rapidly map *Loa loa* (loiasis) using the rapid assessment procedure for loiasis (RAPLOA) method (TDR, 2001; Takougang *et al.* 2002). Individuals with high microfilarial loads of *L. loa* are at increased risk of neurologic serious adverse events following ivermectin treatment against onchocerciasis, and it is therefore important to identify areas of high loiasis risk as part of onchocerciasis control efforts. The RAPLOA approach correlates the proportion of community members reporting a history of eyeworm with the prevalence of high-intensity *L. loa* microfilaremia in that community, and is used to rapidly assess areas at potential risk of treatment-related *L. loa* encephalopathy. Work in Cameroon has demonstrated that RAPLOA and REA could be used simultaneously to rapidly evaluate loiasis and onchocerciasis endemicity in areas targeted by APOC for safe treatment with ivermectin (Wanji *et al.* 2005). As with schistosomiasis, spatial risk mapping can successfully exclude areas of zero or low loiasis transmission prior to rapid assessment surveys (Diggle *et al.* 2007).

The implementation of control efforts against lymphatic filariasis, a disease caused by the filarial parasite *Wuchereria bancrofti*, has also benefited from a rapid mapping approach. WHO recommends that the control of lymphatic filariasis is based on MDA with either ivermectin or diethylcarbamazine (DEC) plus albendazole in areas where prevalence exceeds 1% (WHO, 2000). As a first step in development of national control programmes, comprehensive survey data are essential. Rapid mapping of lymphatic filariasis has been made feasible by the development of immunochromatographic card tests (ICT) for the detection of circulating Og4C3 antigen from adult *W. bancrofti* filarial antigenaemia as well as standardised sampling strategies (WHO, 2000). These sampling strategies have included the use of a spatial sampling grid with either 25 km or 50 km between sampled communities and geostatistical modelling for predicting infection in non-sampled areas, an approach which has been validated (Gyapong and Remme, 2001) and subsequently used to map the distribution of *W. bancrofti* in West Africa (Gyapong *et al.* 2002) and Uganda (Onapa *et al.* 2005). Other analyses suggest, however, that endemic foci can persist within the interstices of the proposed grid and that smaller grids are required (Srividya *et al.* 2002). Another sampling method has been the adaptation of LQAS whereby a minimum of 100 individuals are tested in each community (or school), and then if one or more were found to be positive no further testing was done and MDA will be provided throughout the district in which the community is located. Otherwise, up to 250 individuals are tested to ensure that infection does not exceed 1%. This approach has successfully been used to map infection in Nepal (Sherchand *et al.* 2003) and Haiti (Beau de Rochars *et al.*

2004). LQAS has also been used to monitor coverage after every round of MDA as part of the India national lymphatic filariasis control programme (Vanamail *et al.* 2006).

In addition to lymphatic filariasis and *S. mansoni*, LQAS has been employed to help identify priority areas for interventions targeting blinding trachoma caused by the bacterium *Chlamydia trachomatis* (Myatt *et al.* 2003) and human African trypanosomiasis (Hutin *et al.* 2004). To our knowledge, no attempts have been made thus far to use LQAS for rapid identification of high-risk communities of STH. This is rather surprising, as stool samples can be readily examined for the presence of *S. mansoni* and other helminth eggs, and hence holds promise for prioritizing control not only of intestinal schistosomiasis but also STH (Hugh Sturrock *et al.* manuscript in preparation).

FUTURE DIRECTIONS

In the last two decades, there have been significant developments in methods for identifying at-risk populations for control programmes targeting schistosomiasis, using questionnaire surveys, GIS, and rapid survey methodologies. Practical, low-cost applications of these methods are now being implemented at national and regional scales. More recent interest has focused on the potential of integrating a number of NTD interventions which deliver population-based chemotherapy (WHO, 2006; Hotez *et al.* 2007). The effectiveness of such integrated programmes will crucially depend on the geographical overlap between the different NTDs. However, in spite being co-endemic in individual countries (Brady *et al.* 2006), different NTDs can in certain settings, for example, Uganda (Kolaczinski *et al.* 2007), exhibit limited geographical overlap at sub-national scales, necessitating a more geographically targeted approach for integrated NTD control (Brooker and Utzinger, 2007). Thus, research efforts should be undertaken to determine the optimal strategy of rapidly and simultaneously assessing a number of NTDs. An immediate question that arises is whether it is possible to develop an integrated rapid mapping approach.

As a first step, we have previously proposed that the spatial co-distribution of different NTDs over large geographical areas, such as at continental and regional scales, can be based initially on climate-based disease risk maps (Brooker *et al.* 2006; Brooker, 2007; Brooker and Utzinger, 2007). Such models also have a role in defining co-distribution at local levels (Raso *et al.* 2006b; Brooker and Clements, 2009). At finer scales, however, there will remain a need to undertake rapid and inexpensive assessments of infection levels to guide local integrated control. As outlined here, a number of simple (though not always cheap) approaches, including the use of questionnaires and rapid antigen detection assays, are currently available. An issue which merits further attention, and is indeed germane to the sustainable implementation of NTD control, includes investigation into how these assessment tools can be cost-effectively and practically combined into a single, simple survey methodology.

There are, however, a number of important differences in the mapping approaches used by various NTD programmes, which makes the move to an integrated mapping approach particularly challenging. First, the target age group for sampling differs among the major NTDs: school-aged children for schistosomiasis and STH, persons aged 10 years or above for lymphatic filariasis, and adults for onchocerciasis. Second, the number of individuals to be sampled depends, in part, on the prevalence threshold used to denote the need for MDA: because the threshold for lymphatic filariasis is 1%, up to 250 individuals per community are sampled; this contrasts the 50 children per school for schistosomiasis and STH, or even lower numbers when using LQAS. Third, the inherent spatial heterogeneity of infection varies among the different NTDs (Gyapong *et al.* 2002; Srividya *et al.* 2002; Brooker *et al.* 2004; Raso *et al.* 2006a, b; Brooker, 2007) and this necessarily has implications for the spatial density of sampling. Fourth, various tests and approaches are used to diagnose infection (urine, stool and

blood), and this adds complication and expense to any integrated survey. Finally, the geographical unit at which interventions are implemented varies: whole districts for lymphatic filariasis and perhaps STH, communities or schools for schistosomiasis and onchocerciasis. Any integrated mapping approach would need to reconcile such differences, and the approach would need to be constantly adapted in response to progress made and as a function of the stage of control of different NTDs.

Notwithstanding these challenges, we believe that the issue of rapid, low-cost assessment methods will gain even greater importance as control programmes are increasingly implemented and the aims of control programmes, move from morbidity control to transmission control (Bergquist *et al.* 2009). The identification of remaining pockets of high transmission in areas of low endemicity will become more difficult as a large amount of unnecessary assessments will need to be conducted. Thus, novel approaches to rapid assessment must be developed. A recent study in Brazil has explored the use of schoolchildren as indicators for the identification of *S. mansoni*-positive family members in an area of low endemicity (Massara *et al.* 2006). Although the approach had a relatively low sensitivity (50%), it had a high negative predictive value (87%) suggesting that few positive family members of schoolchildren with a negative stool result are missed. Further evaluation of this approach in other settings and for different diseases is clearly warranted.

For example, as malaria control efforts are intensified in Africa and elsewhere, indeed with renewed efforts towards local elimination and global eradication, there is a pressing need for rapid assessment methods to guide interventions. The usefulness of rapid malaria assessment in urban areas, including collection of health statistics, health facility-based surveys, vector breeding site surveys, and school parasite surveys, has recently been evaluated in four cities across Africa (Wang *et al.* 2005). It would be interesting to evaluate this approach and to investigate alternative rapid malaria assessment methods, including LQAS (Rabarijaona *et al.* 2001), in rural settings.

The research reviewed here provide salient, evidence-based approaches to rapidly understanding the prevalence and distribution of schistosomiasis and other NTDs in order to facilitate the design and implementation of sustainable development programmes. Alternative rapid assessment methods and their cost-effectiveness should continue to be investigated. We hope this review will stimulate a concerted research effort in the future.

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References

- Ansell J, Guyatt HL. Comparative cost-effectiveness of diagnostic tests for urinary schistosomiasis and the implications for school health programmes. *Annals of Tropical Medicine and Parasitology* 2002;96:145–153. [PubMed: 12080975]
- Ansell J, Guyatt H, Hall A, Kihamia C, Bundy DAP. The effects of sex and age of responders on the reliability of self-diagnosed infection: a study of self-reported urinary schistosomiasis in Tanzanian school children. *Social Science and Medicine* 2001;53:957–967. [PubMed: 11522140]
- Ayele B, Erko B, Legesse M, Hailu A, Medhin G. Evaluation of circulating cathodic antigen (CCA) strip for diagnosis of urinary schistosomiasis in Hassoba school children, Afar, Ethiopia. *Parasite* 2008;15:69–75. [PubMed: 18416249]

- Ayong LS, Tume CB, Wembe FE, Simo G, Asonganyi T, Lando G, Ngu JL. Development and evaluation of an antigen detection dipstick assay for the diagnosis of human onchocerciasis. *Tropical Medicine and International Health* 2005;10:228–233. [PubMed: 15730506]
- Beau de Rochars MV, Milord MD, St Jean Y, Désormeaux AM, Dorvil JJ, Lafontant JG, Addiss DG, Streit TG. Geographic distribution of lymphatic filariasis in Haiti. *American Journal of Tropical Medicine and Hygiene* 2004;71:598–601. [PubMed: 15569791]
- Bergquist R, Johansen MV, Utzinger J. Diagnostic dilemmas in helminthology: what tools to use and when? *Trends in Parasitology* 2009;25 in press
- Birrie H, Medhin G, Jemaneh L. Comparison of urine filtration and a chemical reagent strip in the diagnosis of urinary schistosomiasis in Ethiopia. *East African Medical Journal* 1995;72:180–185. [PubMed: 7796771]
- Brady MA, Hooper PJ, Ottesen EA. Projected benefits from integrating NTD programs in sub-Saharan Africa. *Trends in Parasitology* 2006;22:285–291. [PubMed: 16730230]
- Brooker S. Spatial epidemiology of human schistosomiasis in Africa: risk models, transmission dynamics and control. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 2007;101:1–8. [PubMed: 17055547]
- Brooker S, Clements ACA. Spatial heterogeneity of parasite co-infection: determinants and geostatistical prediction at regional scales. *International Journal for Parasitology* 2009;39:591–597. [PubMed: 19073189]
- Brooker S, Clements ACA, Bundy DAP. Global epidemiology, ecology and control of soil-transmitted helminth infections. *Advances in Parasitology* 2006;62:223–265.
- Brooker S, Kabatereine NB, Fleming F, Devlin N. Cost and cost-effectiveness of nationwide school-based helminth control in Uganda: intra-country variation and effects of scaling-up. *Health Policy and Planning* 2008;23:24–35. [PubMed: 18024966]
- Brooker S, Kabatereine NB, Myatt M, Stothard JR, Fenwick A. Rapid assessment of *Schistosoma mansoni*: the validity, applicability and cost-effectiveness of the lot quality assurance sampling method in Uganda. *Tropical Medicine and International Health* 2005;10:647–658. [PubMed: 15960703]
- Brooker S, Kabatereine NB, Tukahebwa EM, Kazibwe F. Spatial analysis of the distribution of intestinal nematode infections in Uganda. *Epidemiology and Infection* 2004;132:1065–1071. [PubMed: 15635963]
- Brooker S, Miguel EA, Waswa P, Namunyu R, Moulin S, Guyatt H, Bundy DAP. The potential of rapid screening methods for *Schistosoma mansoni* in western Kenya. *Annals of Tropical Medicine and Parasitology* 2001;95:343–351. [PubMed: 11454244]
- Brooker S, Rowlands M, Haller L, Savioli L, Bundy DAP. Towards an atlas of human helminth infection in sub-Saharan Africa: the use of geographical information systems (GIS). *Parasitology Today* 2000;16:303–307. [PubMed: 10858650]
- Brooker S, Utzinger J. Integrated disease mapping in a polyparasitic world. *Geospatial Health* 2007;1:141–146. [PubMed: 18686239]
- Bundy, DAP.; Shaeffer, S.; Jukes, M.; Beegle, K.; Gillespie, A.; Drake, L.; Lee, Seung-hee Frances; Hoffman, AM.; Jones, J.; Mitchell, A.; Wright, C.; Barcelona, D.; Camara, B.; Golmar, C.; Savioli, L.; Takeuchi, T.; Sembene, M. School based health and nutrition programs. In: Jamison, D.; Breman, JG.; Measham, AR.; Alleyne, G.; Claeson, M.; Evans, D.; Jha, P.; Mills, A.; Musgrove, P., editors. *Disease Control Priorities in Developing Countries. Vol. Second Edition. The World Bank and Oxford University Press; NY, USA: 2006. p. 1091-1108.*
- Chappuis F, Rijal S, Jha UK, Desjeux P, Karki BM, Koirala S, Loutan L, Boelaert M. Field validity, reproducibility and feasibility of diagnostic tests for visceral leishmaniasis in rural Nepal. *Tropical Medicine and International Health* 2006;11:31–40. [PubMed: 16398753]
- Clements ACA, Barnett AG, Nyandindi U, Lwambo NJ, Kihamia CM, Blair L. Age and gender effects in self-reported urinary schistosomiasis in Tanzania. *Tropical Medicine and International Health* 2008a;13:713–721. [PubMed: 18419586]
- Clements ACA, Brooker S, Nyandindi U, Fenwick A, Blair L. Bayesian spatial analysis of a national urinary schistosomiasis questionnaire to assist geographic targeting of schistosomiasis control in

- Tanzania, East Africa. *International Journal for Parasitology* 2008b;38:401–415. [PubMed: 17920605]
- Danso-Appiah A, Garner P, Olliaro PL, Utzinger J. Treatment of urinary schistosomiasis: methodological issues and research needs identified through a Cochrane systematic review. *Parasitology*. (to be published in the same special issue of *Parasitology*: COPY EDITOR, please provide volume and page numbers). 2009
- de Vlas SJ, Gryseels B. Underestimation of *Schistosoma mansoni* prevalences. *Parasitology Today* 1992;8:274–277. [PubMed: 15463638]
- Diggle PJ, Thomson MC, Christensen OF, Rowlingson B, Obsomer V, Gardon J, Wanji S, Takougang I, Enyong P, Kamgno J, Remme JH, Boussinesq M, Molyneux DH. Spatial modelling and the prediction of *Loa loa* risk: decision making under uncertainty. *Annals of Tropical Medicine and Parasitology* 2007;101:499–509. [PubMed: 17716433]
- Doumenge, JP.; Mott, KE.; Cheung, C.; Villenave, D.; Chapuis, O.; Perrin, MF.; Read-Thomas, G. Atlas of the global distribution of schistosomiasis. World Health Organization; Geneva: 1987.
- Engels D, Sinzinkayo E, Gryseels B. Day-to-day egg count fluctuation in *Schistosoma mansoni* infection and its operational implications. *American Journal of Tropical Medicine and Hygiene* 1996;54:319–324. [PubMed: 8615440]
- Engels D, Sinzinkayo E, Gryseels B. Intraspecimen fecal egg count variation in *Schistosoma mansoni* infection. *American Journal of Tropical Medicine and Hygiene* 1997;57:571–577. [PubMed: 9392598]
- French MD, Rollinson D, Basáñez MG, Mgeni AF, Khamis IS, Stothard JR. School-based control of urinary schistosomiasis on Zanzibar, Tanzania: monitoring micro-haematuria with reagent strips as a rapid urological assessment. *Journal of Pediatric Urology* 2007;3:364–368. [PubMed: 18947774]
- Gryseels B. Morbidity due to infection with *Schistosoma mansoni*: an update. *Tropical and Geographical Medicine* 1992;44:189–200. [PubMed: 1455521]
- Gyapong JO, Kyelem D, Kleinschmidt I, Agbo K, Ahouandogbo F, Gaba J, Owusu-Banahene G, Sanou S, Sodahlon YK, Biswas G, Kale OO, Molyneux DH, Roungou JB, Thomson MC, Remme J. The use of spatial analysis in mapping the distribution of bancroftian filariasis in four West African countries. *Annals of Tropical Medicine and Parasitology* 2002;96:695–705. [PubMed: 12537631]
- Gyapong JO, Remme JFH. Validation of a rapid assessment method for the distribution of Bancroftian filariasis. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 2001;95:681–686. [PubMed: 11816445]
- Hall A, Fentiman A. Blood in the urine of adolescent girls in an area of Ghana with a low prevalence of infection with *Schistosoma haematobium*. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 1999;93:411–412. [PubMed: 10674091]
- Hotez PJ, Molyneux DH, Fenwick A, Kumaresan J, Ehrlich Sachs S, Sachs JD, Savioli L. Control of neglected tropical diseases. *New England Journal of Medicine* 2007;357:1018–1027. [PubMed: 17804846]
- Hutin YJ, Legros D, Owini V, Brown V, Lee E, Mbulamberi D, Paquet C. *Trypanosoma brucei gambiense* trypanosomiasis in Terego county, northern Uganda, 1996: a lot quality assurance sampling survey. *American Journal of Tropical Medicine and Hygiene* 2004;70:390–394. [PubMed: 15100452]
- Kabatereine NB, Brooker S, Koukounari A, Kazibwe F, Tukahebwa EM, Fleming FM, Zhang Y, Webster JP, Stothard JR, Fenwick A. Impact of a national helminth control programme on infection and morbidity in Ugandan schoolchildren. *Bulletin of the World Health Organization* 2007;85:91–99. [PubMed: 17308729]
- Katarwa M, Onapa AW, Nakileza B. Rapid epidemiological mapping of onchocerciasis in areas of Uganda where *Simulium neavei* sl is the vector. *East African Medical Journal* 1999;76:440–446. [PubMed: 10520349]
- Kolaczinski J, Kabatereine NB, Onapa A, Ndyomugyengi R, Kakmebo AS, Brooker S. Neglected tropical diseases in Uganda: the prospect and challenge of integrated control. *Trends in Parasitology* 2007;23:485–493. [PubMed: 17826335]
- Kurowski C, Wyss K, Abdulla S, Mills A. Scaling up priority health interventions in Tanzania: the human resources challenge. *Health Policy and Planning* 2007;22:113–127. [PubMed: 17485419]

- Lammie PJ, Fenwick A, Utzinger J. A blueprint for success: integration of neglected tropical disease control programmes. *Trends in Parasitology* 2006;22:313–321. [PubMed: 16713738]
- Legesse M, Erko B. Field-based evaluation of a reagent strip test for diagnosis of *Schistosoma mansoni* by detecting circulating cathodic antigen in urine before and after chemotherapy. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 2007;101:668–673. [PubMed: 17368699]
- Legesse M, Erko B. Field-based evaluation of a reagent strip test for diagnosis of schistosomiasis mansoni by detecting circulating cathodic antigen (CCA) in urine in low endemic area in Ethiopia. *Parasite* 2008;15:151–155. [PubMed: 18642508]
- Lengeler C, de Savigny D, Mshinda H, Mayombana C, Tayari S, Hatz C, Degrémont A, Tanner M. Community-based questionnaires and health statistics as tools for the cost-efficient identification of communities at risk of urinary schistosomiasis. *International Journal of Epidemiology* 1991a;20:796–807. [PubMed: 1955266]
- Lengeler C, Kilima P, Mshinda H, Morona D, Hatz C, Tanner M. Rapid, low-cost, two-step method to screen for urinary schistosomiasis at the district level: the Kilosa experience. *Bulletin of the World Health Organization* 1991b;69:179–189. [PubMed: 1907220]
- Lengeler C, Utzinger J, Tanner M. Questionnaires for rapid screening of schistosomiasis in sub-Saharan Africa. *Bulletin of the World Health Organization* 2002;80:235–242. [PubMed: 11984610]
- Mafe MA. The diagnostic potential of three indirect tests for urinary schistosomiasis in Nigeria. *Acta Tropica* 1997;68:277–284. [PubMed: 9492912]
- Mafe MA, von Stamm T, Utzinger J, N'Goran EK. Control of urinary schistosomiasis: an investigation into the effective use of questionnaires to identify high-risk communities and individuals in Niger State, Nigeria. *Tropical Medicine and International Health* 2000;5:53–63. [PubMed: 10672206]
- Marx A, Pewsner D, Egger M, Nüesch R, Bucher HC, Genton B, Hatz C, Jüni P. Meta-analysis: accuracy of rapid tests for malaria in travelers returning from endemic areas. *Annals of Internal Medicine* 2005;142:836–846. [PubMed: 15897534]
- Massara CL, Peixoto SV, Enk MJ, da Silva Barros H, dos Santos Carvalho O, Sakurai E, Schall V. Evaluation of an improved approach using residences of schistosomiasis-positive school children to identify carriers in an area of low endemicity. *American Journal of Tropical Medicine and Hygiene* 2006;74:495–499. [PubMed: 16525112]
- Mathieu E, Deming M, Lammie PJ, McLaughlin SI, Beach MJ, Deodat DJ, Addiss DG. Comparison of methods for estimating drug coverage for filariasis elimination, Leogane commune, Haiti. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 2003;97:501–505. [PubMed: 15307410]
- Molyneux DH, Hotez PJ, Fenwick A. “Rapid-impact interventions”: how a policy of integrated control for Africa's neglected tropical diseases could benefit the poor. *PLoS Medicine* 2005;2:e336. [PubMed: 16212468]
- Moody A. Rapid diagnostic tests for malaria parasites. *Clinical Microbiology Reviews* 2002;15:66–78. [PubMed: 11781267]
- Mtasiwa D, Mayombana C, Kilima P, Tanner M. Validation of reagent sticks in diagnosing urinary schistosomiasis in an urban setting. *East African Medical Journal* 1996;73:198–200. [PubMed: 8698022]
- Myatt M, Limburg H, Minassian D, Katyola D. Field trial of applicability of lot quality assurance sampling survey method for rapid assessment of prevalence of active trachoma. *Bulletin of the World Health Organization* 2003;81:877–885. [PubMed: 14997240]
- N'Goran EK, Utzinger J, Traore M, Lengeler C, Tanner M. Identification rapide par questionnaire des principaux foyers de bilharziose urinaire au centre de la Côte d'Ivoire. *Médecine Tropicale* 1998;58:253–260.
- Ngoumou P, Walsh JF, Mace JM. A rapid mapping technique for the prevalence and distribution of onchocerciasis: a Cameroon case study. *Annals of Tropical Medicine and Parasitology* 1994;88:463–474. [PubMed: 7979636]
- N'Guessan NA, Acka CA, Utzinger J, N'Goran EK. Identification des régions à haut risque de schistosomoses en Côte d'Ivoire. *Bulletin de la Société de Pathologie Exotique* 2007;100:119–123.

- Noma M, Nwoke BE, Nutall I, Tambala PA, Enyong P, Namsenmo A, Remme J, Amazigo UV, Kale OO, Sékétéli A. Rapid epidemiological mapping of onchocerciasis (REMO): its application by the African programme for onchocerciasis control (APOC). *Annals of Tropical Medicine and Parasitology* 2002;96:S29–S39. [PubMed: 12081248]
- Onapa AW, Simonsen PE, Baehr I, Pedersen EM. Rapid assessment of the geographical distribution of lymphatic filariasis in Uganda, by screening of schoolchildren for circulating filarial antigens. *Annals in Tropical Medicine and Parasitology* 2005;99:141–153.
- Partnership for Child Development. Better health, nutrition and education for the school aged child. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 1997;91:1–2. [PubMed: 9093613]
- Poggensee G, Krantz I, Kiwelu I, Feldmeier H. Screening of Tanzanian women of childbearing age for urinary schistosomiasis: validity of urine reagent strip readings and self-reported symptoms. *Bulletin of the World Health Organization* 2000;78:542–548. [PubMed: 10885183]
- Rabarijaona L, Rakotomanana F, Ranaivo L, Raharimalala L, Modiano D, Boisier P, De Giorgi F, Raveloson N, Jambou R. Validity of lot quality assurance sampling to optimize falciparum malaria surveys in low-transmission areas. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 2001;95:267–269. [PubMed: 11490993]
- Raso G, Vounatsou P, Gosoni L, Tanner M, N'Goran EK, Utzinger J. Risk factors and spatial patterns of hookworm infection among schoolchildren in a rural area of western Côte d'Ivoire. *International Journal for Parasitology* 2006a;36:201–210. [PubMed: 16259987]
- Raso G, Vounatsou P, McManus DP, Utzinger J. Bayesian risk maps for *Schistosoma mansoni* and hookworm mono-infections in a setting where both parasites co-exist. *Geospatial Health* 2007;2:85–96. [PubMed: 18686258]
- Raso, G.; Vounatsou, P.; Singer, BH.; N'Goran, EK.; Tanner, M.; Utzinger, J. An integrated approach for risk profiling and spatial prediction of *Schistosoma mansoni*-hookworm coinfection; *Proceedings of the National Academy of Sciences USA*; 2006b. p. 6934–6939.
- Robertson SE, Valadez JJ. Global review of health care surveys using lot quality assurance sampling (LQAS), 1984–2004. *Social Science and Medicine* 2006;63:1648–1660. [PubMed: 16764978]
- Sherchand JB, Obsomer V, Thakur GD, Hommel M. Mapping of lymphatic filariasis in Nepal. *Filaria Journal* 2003;2:7. [PubMed: 12694630]
- Simoonga C, Utzinger J, Brooker S, Vounatsou P, Appleton CC, Stensgaard AS, Olsen A, Kristensen TK. Remote sensing, geographical information system and spatial analysis for schistosomiasis epidemiology and ecology in Africa. *Parasitology*. 2009(to be published in the same special issue of *Parasitology*: COPY EDITOR, please provide volume and page numbers)
- Srividya A, Michael E, Palaniyandi M, Pani SP, Das PK. A geostatistical analysis of the geographic distribution of lymphatic filariasis prevalence in southern India. *American Journal of Tropical Medicine and Hygiene* 2002;67:480–489. [PubMed: 12479548]
- Stothard JR, Kabatereine NB, Tukahebwa EM, Kazibwe F, Rollinson D, Mathieson W, Webster JP, Fenwick A. Use of circulating cathodic antigen (CCA) dipsticks for detection of intestinal and urinary schistosomiasis. *Acta Tropica* 2006;97:219–228. [PubMed: 16386231]
- Takougang I, Meremikwu M, Wandji S, Yenshu EV, Aripko B, Lamlenn SB, Eka BL, Enyong P, Meli J, Kale O, Remme JH. Rapid assessment method for prevalence and intensity of *Loa loa* infection. *Bulletin of the World Health Organization* 2002;80:852–858. [PubMed: 12481206]
- TDR. Rapid assessment procedures for loiasis: report of a multi-centre study. *UNDP/World Bank/WHO Special Programme for Research and Training in Tropical Diseases*; Geneva: 2001.
- Utzinger J, Bergquist R, Xiao SH, Singer BH, Tanner M. Sustainable schistosomiasis control – the way forward. *Lancet* 2003;362:1932–1934. [PubMed: 14667754]
- Utzinger J, Booth M, N'Goran EK, Müller I, Tanner M, Lengeler C. Relative contribution of day-to-day and intra-specimen variation in faecal egg counts of *Schistosoma mansoni* before and after treatment with praziquantel. *Parasitology* 2001;122:537–544. [PubMed: 11393827]
- Utzinger J, de Savigny D. Control of neglected tropical diseases: integrated chemotherapy and beyond. *PLoS Medicine* 2006;3:e112. [PubMed: 16435910]
- Utzinger J, N'Goran EK, Esse Aya CM, Acka Adjoua C, Lohourignon KL, Tanner M, Lengeler C. *Schistosoma mansoni*, intestinal parasites and perceived morbidity indicators in schoolchildren in a

- rural endemic area of western Côte d'Ivoire. *Tropical Medicine and International Health* 1998;3:711–720. [PubMed: 9754666]
- Utzinger J, N'Goran EK, Ossey YA, Booth M, Traoré M, Lohourignon KL, Allangba A, Ahiba LA, Tanner M, Lengeler C. Rapid screening for *Schistosoma mansoni* in western Côte d'Ivoire using a simple school questionnaire. *Bulletin of the World Health Organization* 2000;78:389–398. [PubMed: 10812739]
- van Dam GJ, Wichers JH, Falcao Ferreira TM, Ghati D, van Amerongen A, Deelder AM. Diagnosis of schistosomiasis by reagent strip test for detection of circulating cathodic antigen. *Journal of Clinical Microbiology* 2004;42:5458–61. [PubMed: 15583265]
- van der Werf MJ, Borsboom GJ, de Vlas SJ. No effect of recall period length on prevalence of self-reported haematuria in *Schistosoma haematobium*-endemic areas. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 2003;97:373–374. [PubMed: 15259459]
- van Etten L, van Lieshout L, Mansour MM, Deelder AM. A reagent strip antigen capture assay for the assessment of cure of schistosomiasis patients. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 1997;91:154–155. [PubMed: 9196755]
- Vanamail P, Subramanian S, Srividya A, Ravi R, Krishnamoorthy K, Das PK. Operational feasibility of lot quality assurance sampling (LQAS) as a tool in routine process monitoring of filariasis control programmes. *Tropical Medicine and International Health* 2006;11:1256–1263. [PubMed: 16903888]
- Vounatsou P, Raso G, Tanner M, N'Goran EK, Utzinger J. Bayesian geostatistical modelling for risk profiling of schistosomiasis: a review. *Parasitology*. 2009(to be published in the same special issue of Parasitology: COPY EDITOR, please provide volume and page numbers)
- Wang SJ, Lengeler C, Smith TA, Vounatsou P, Cissé G, Diallo DA, Akogbeto M, Mtasiwa D, Teklehaimanot A, Tanner M. Rapid urban malaria appraisal (RUMA) in sub-Saharan Africa. *Malaria Journal* 2005;4:40. [PubMed: 16153298]
- Wanji S, Tendongfor N, Esum M, Yundze SSJ, Taylor MJ, Enyong P. Combined utilisation of rapid assessment procedures for loiasis (RAPLOA) and onchocerciasis (REA) in rain forest villages of Cameroon. *Filaria Journal* 2005;4:2. [PubMed: 15817124]
- Weil GJ, Lammie PJ, Weiss N. The ICT filariasis test: a rapid-format antigen test for diagnosis of bancroftian filariasis. *Parasitology Today* 1997;13:401–404. [PubMed: 15275155]
- WHO. The schistosomiasis manual. World Health Organization; Geneva: 1995. Social and Economic Research Project Reports No. 3
- WHO. Operational guidelines for rapid mapping of Bancroftian filariasis in Africa. World Health Organization; Geneva: 2000. WHO/CDS/CPE/CEE/2000.9
- WHO. Prevention and control of schistosomiasis and soil-transmitted helminthiasis: report of a WHO expert committee. 2002. p. 1-57. WHO Technical Report Series No. 912
- WHO. Preventive chemotherapy in human helminthiasis: coordinated use of anthelmintic drugs in control interventions: a manual for health professionals and programme managers. World Health Organization; Geneva: 2006.

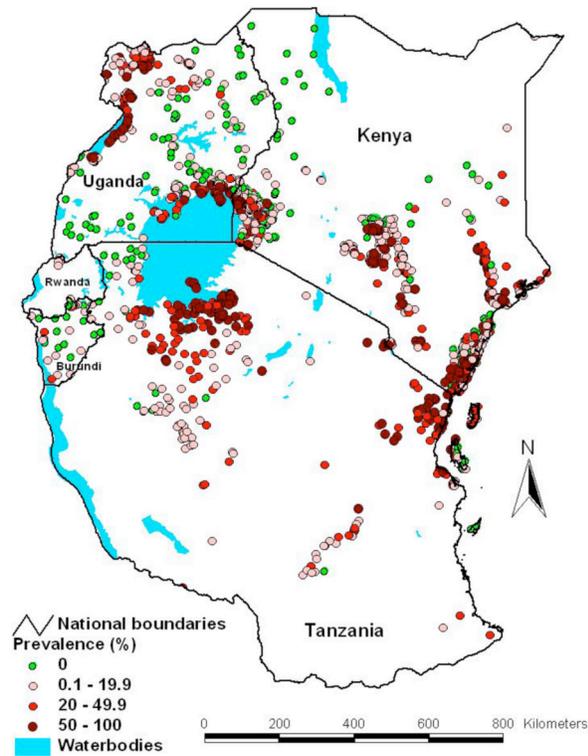


Fig. 1.

The distribution of schistosomiasis in East and Southern Africa based on current progress in updating the Global Atlas of Human Helminth Infection (for further details see Brooker *et al.* 2000). The data included 2,498 survey locations reporting either *S. mansoni* or *S. haematobium*, and the highest prevalence of either species is presented. Information is based on electronic searches of formal literature and grey literature databases, as well as using personal contacts with schistosomiasis research scientists and control personnel. In the searches strict inclusion criteria are employed: only random or complete community-sample surveys conducted post-1980, where parasite species and age groups are defined. Surveys are geopositioned using Microsoft Encarta encyclopaedia and electronic gazetteer sources along with personal communication with authors.

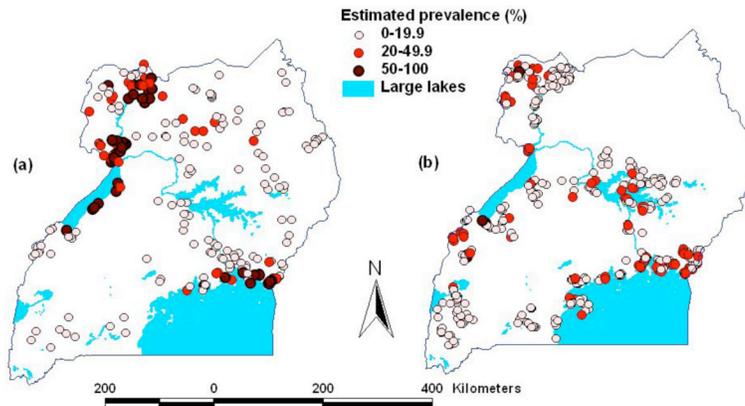


Fig. 2.

Estimated distribution of *S. mansoni* in Uganda pre-2004 and in 2006. The pre-2004 data were collected from 201 schools and based on stool examination using the Kato-Katz method (Kabaterine *et al.* 2004). The 2006 data were collected from 429 schools in 31 districts across the country and based on a rapid mapping survey using LQAS technique (Brooker *et al.* 2005). Here, four teams, comprised of one supervisor, two technicians and a driver visited different regions of the country and sampled eight schools per day over 3 days per district. Geographical coordinates, using hand-held global positioning system (GPS) devices, were also recorded for each school which allowed, once all the data had been collected, the creation of prevalence maps using GIS software.

Table 1

Diagnostic performance of reagent strip testing for microhaematuria compared to urine filtration (considered as the diagnostic 'gold' standard), performed by research teams in different African settings

Setting, country (year)	No. of individuals	Diagnostic performance of reagent strips ^a				Reference	
		Examined	<i>S. haematobium</i> positive (%)	Sensitivity	Specificity		Positive predictive value
Itala district, Dar es Salaam, Tanzania (1992)	404	273 (67.6)	93 (89-95)	86 (79-91)	93 (90-96)	85 (78-90)	Mtasiwa <i>et al.</i> (1996)
4 villages, Kainji Lake area, Nigeria (not known)	1065	606 (57.4)	69 (65-72)	80 (76-83.3)	82 (78-85)	65 (61-69)	Mafe (1997)
Borgu local government, Nigeria (1998)	529	172 (32.5)	78 (71-84)	83 (79-87)	69 (62-75)	89 (85-92)	Mafe <i>et al.</i> (2000)
Enta Doyta, Awash River valley, Ethiopia (1989)	226	69 (30.5)	78 (66-87) ^b	67 (59-74) ^b	51 (41-61) ^b	88 (80-83) ^b	Birnie <i>et al.</i> (1995)
Dahitele, Awash River valley, Ethiopia (1989)	124	26 (21.0)	77 (56-90) ^b	83 (73-89) ^b	54 (37-70) ^b	93 (85-97) ^b	Birnie <i>et al.</i> (1995)
24 schools Unguja Island, Zanzibar, Tanzania (2004)	Boys: 892 Girls: 1084	131 (14.7) 129 (11.9)	91 (89-93) 77 (74-80)	98 (97-99) 97 (96-98)	88 (86-90) 76 (73-78)	98 (98-99) 97 (96-98)	French <i>et al.</i> (2007)
24 schools Unguja Island, Zanzibar, Tanzania (2005)	Boys: 1525 Girls: 1747	228 (14.9) 176 (10.1)	91 (89-92) 91 (90-92)	98 (98-99) 98 (97-98)	90 (89-92) 81 (79-83)	98 (98-99) 99 (99-100)	French <i>et al.</i> (2007)
24 schools Unguja Island, Zanzibar, Tanzania (2006)	Boys: 1521 Girls: 1784	154 (10.1) 105 (5.9)	86 (86-88) 84 (82-86)	99 (98-100) 98 (98-99)	90 (89-91) 77 (75-79)	98 (98-99) 99 (99-100)	French <i>et al.</i> (2007)
5 districts, south-central Côte d'Ivoire (1996)	1336	153 (11.5)	67 (59-74)	97 (95-98)	71 (63-78)	96 (94-97)	N'Goran <i>et al.</i> (1998)
Afambo, Awash River valley, Ethiopia (1989)	155	5 (3.2)	80 (30-99) ^b	87 (80-92) ^b	17 (6-38) ^b	99 (95-100) ^b	Birnie <i>et al.</i> (1995)

^a Values in brackets indicate 95% confidence intervals

^b Values have been recalculated, including 95% confidence intervals.

Table 2

Diagnostic performance of reagent strip testing for microhaematuria carried out by previously trained teachers compared to research teams in different African settings (the results from the research teams were considered as the diagnostic 'gold' standard)

Setting, country (year)	No. of individuals		Diagnostic performance of reagent strips ^a				Reference
	Examined	<i>S. haematobium</i> positive (%)	Sensitivity	Specificity	Positive predictive value	Negative predictive value	
Borgu local government, Nigeria (1998)	486	177 (36.4)	83 (76-88)	89 (85-92)	81 (75-86)	90 (86-93)	Mafe <i>et al.</i> (2000)
Djékanou, Kokumbo, Iaafo, Triassalé and Toumodi, Côte d'Ivoire (1996)	1336	138 (10.3)	71 (63-78)	96 (95-97)	69 (60-76)	97 (95-98)	N'Goran <i>et al.</i> (1998)

^a Values in brackets indicate 95% confidence intervals