

Published in final edited form as:

*Parasitology*. 2006 December ; 133(Pt 6): 711–719.

## Bayesian geostatistical prediction of the intensity of infection with *Schistosoma mansoni* in East Africa

ARCHIE C.A. CLEMENTS<sup>1,2</sup>, RANA MOYEED<sup>3</sup>, and SIMON BROOKER<sup>1</sup>

<sup>1</sup> Department of Infectious and Tropical Diseases, London School of Hygiene and Tropical Medicine, London, UK

<sup>2</sup> Schistosomiasis Control Initiative, Department of Infectious Disease Epidemiology, Imperial College London, London, UK

<sup>3</sup> School of Mathematics and Statistics, University of Plymouth, Plymouth, UK

### SUMMARY

A Bayesian geostatistical model was developed to predict the intensity of infection with *Schistosoma mansoni* in East Africa. Epidemiological data from purposively-designed and standardized surveys were available for 31,458 schoolchildren (90% aged between 6-16 years) from 459 locations across the region and used in combination with remote sensing environmental data to identify factors associated with spatial variation in infection patterns. The geostatistical model explicitly takes into account the highly aggregated distribution of parasite distribution by fitting a negative binomial distribution to the data and accounts for spatial correlation. Results identify the role of environmental risk factors in explaining geographical heterogeneity in infection intensity and show how these factors can be used to develop a predictive map. Such a map has important implications for schistosomiasis control programmes in the region.

### Keywords

Bayesian models; geostatistical prediction; negative binomial distribution; *Schistosoma mansoni*; schistosomiasis; East Africa

### INTRODUCTION

It is well recognised that reliable maps of the spatial distribution of parasitic diseases are required to help plan and evaluate disease control, and during the last decade there has been growing research interest in modelling disease risk (Hay et al., 2000, 2006). Most studies have modelled point prevalence data (Thomson et al., 1999; Kleinschmidt et al. 2001; Brooker et al., 2006; Clements et al., 2006) since this is an easily collected and readily available indicator. However, although prevalence data are often used for making recommendations for the need for control, data on the intensity of infection has greater relevance to the transmission dynamics of a given host-parasite system and for understanding the occurrence of morbidity (Anderson and May, 1991).

The modelling of infection intensity is nonetheless complicated by the fact that parasite distributions are highly aggregated within communities (Woolhouse et al., 1998; Shaw et al., 1998; Basáñez and Boussinesq, 1999) and by the presence of spatial correlation. In addition to parasite distributions, aggregated - or so-called overdispersed - distributions are a

characteristic of several epidemiological phenomena including social networks (Zheng et al., 2006), infection exposure (Woolhouse et al., 1998) and wildlife distributions (Thogmartin et al., 2004). Aggregated distributions can often be fitted by the negative binomial distribution (Anderson, 1993) and previous analysis has employed negative binomial regression modelling to investigate the relationship between intensity of parasitic infection and a range of individual- and group-level risk factors (Nodtvedt et al., 2002; Yapi et al., 2005).

Studies modelling the spatial distribution of parasitic diseases have increasingly adopted a fully Bayesian geostatistical approach (Carabin et al., 2003; Clements et al., 2006; Diggle et al., 2002; Gemperli et al., 2004, 2006; Raso et al., 2005, 2006; Yang et al., 2005). This approach explicitly incorporates the spatial correlation structure of the data, the ability to include covariate effects and the comprehensive representation of uncertainty in the model outputs. Recently, Alexander and others (Alexander et al., 2000, 2003) have presented a negative binomial spatial model, which incorporates spatial correlation and adjusts for individual-level covariates such as age and sex. Their analysis however was explicative rather than predictive (limiting its utility for disease control decision-making), conducted on a small geographical scale and, unlike the Bayesian geostatistical modelling studies described above, did not include environmental factors which may have helped explain some of the observed heterogeneity.

In the present study, we extend the approach of Alexander and colleagues to model the spatial distribution of the intensity of *Schistosoma mansoni* infection in East Africa. The modelling of intensity of infection, as measured by quantitative faecal egg counts, has particular relevance for the targeting of control activities since intensity is related to schistosome transmission dynamics and is a key determinant of schistosome-related morbidity with high levels likely to induce greater morbidity. Our adopted approach provides estimates of the effects of various factors influencing infection patterns including individual-level variables, such as age and sex, and location-level environmental variables. We then follow a Bayesian geostatistical approach to generate predictive maps of infection intensity across the region, and assess uncertainty associated with predictions.

## PATIENTS AND METHODS

### Data sources

Individual-level data on intensity of *Schistosoma mansoni* infection were obtained from cross-sectional random samples of schoolchildren from dedicated school surveys conducted between 1999 and 2004 at 459 locations by national research teams under the auspices of the Schistosomiasis Control Initiative (SCI) in Tanzania (Clements et al., 2006) and in Uganda (Kabaterine et al., 2004) and by research projects in western Kenya (Brooker et al., 2001; Clarke et al., 2005). The locations of surveyed schools are shown in Figure 1 and details of the studies are shown in Table 1. These data cover a large contiguous area of East Africa covering an area of approximately 1,260 km (North/South)  $\times$  590 km (East/West) and, to our knowledge, represent some of the most comprehensive geo-referenced data on the intensity of infection for any parasitic disease. No previous mass treatment had been undertaken in any of the schools. The details of the survey design, data collection and acquisition of ethical approval are reported in the original publications. Intensity of infection was based on the microscopic examination of two Kato-Katz smears derived from a single stool specimen, and expressed as eggs per gram of faeces (epg). As well as information on quantitative egg counts, individual-level factors including age and sex were also included in the final dataset.

Long-term climatic data (1982-2000) were obtained from the National Oceanographic and Atmospheric Administration's (NOAA) Advanced Very High Radiometer (AVHRR) which provides indirect estimates of land surface temperature (LST) and normalized difference vegetation index (NDVI). Estimates of elevation were obtained from an interpolated digital

elevation model from the Global Land Information System (GLIS) of the United States Geological Survey (<http://edcwww.cr.usgs.gov/landdaac/gtopo30/>) and the location of inland water-bodies, obtained from the US National Imagery and Mapping Agency (NIMA) and transformed by FAO/GIS at the Food and Agriculture Organisation (FAO). For each school, Euclidian distance to in-land water bodies was calculated. The degree of urbanisation, categorised into urban, periurban, rural and extreme rural, was derived from Tatem and other (Tatem et al., 2005). These data were imported into the GIS ArcMap 8.1 (ESRI, Redlands, CA) and linked by geographic location to the parasitological data.

### Statistical analysis

Initial analysis indicated that the distribution of egg counts were not Poisson distributed, with a predominance of zero and low values, such that the median egg count was 0 eggs/gram and the variance was  $3.6 \times 10^5$ , clearly indicative of overdispersion. This justified a multi-variable regression approach (to account for some of the additional sources of heterogeneity) using the negative binomial distribution (to model the overdispersion) in our statistical modelling and prediction. Negative binomial regression models were fitted in Stata/SE version 8.1 (Stata Corporation, College Station, Texas). Initially, univariate analyses were conducted and variables with Wald's  $P > 0.2$  were excluded from further analyses. With the remaining variables, backwards-stepwise regression was conducted using Wald's  $P > 0.1$  as the exit criterion and  $P \leq 0.05$  as the entry criterion. In the final model, sex was included as an individual-level covariate and elevation and distance to perennial water bodies were included as school-level covariates. Modelling either the overdispersion parameter as a function of a range of covariates or the zero values separately from the positive values (using a zero-inflated model) did not improve the overall fit, justifying the subsequent use of a standard negative binomial regression model with a fixed value for the overdispersion parameter.

To provide an explanation of the overdispersion in the data and in particular to assess whether the overdispersion is spatially structured, we followed the approach of Alexander et al. (2000). In this, the total egg count of each individual was modelled as a negative binomial variate with overdispersion parameter  $k > 0$  which incorporates extra-Poisson variation. Larger values of  $k$  indicate less variability, with the limiting case  $k = \infty$  corresponding to the Poisson distribution. We model the logarithm of the mean of the distribution as an additive function of the individual-level covariate sex, the two school-level covariates elevation and distance to nearest inland perennial water body and a spatially-structured school-level random-effect. The spatial random-effect was modelled as a stationary Gaussian process with mean 0, variance  $\sigma^2$  and correlation function  $\exp(-d_{ij}/\alpha)$ , where  $d_{ij}$  is the distance between villages  $i$  and  $j$  and the parameter  $\alpha$  measures the rate at which the spatial correlation decays over distance, with  $\alpha \log 2$  being a characteristic length, which we call the 'half-distance', over which the correlation reduces by half, and  $3\alpha$  being the distance at which the correlation reduces to 0.05.

Bayesian inference was implemented via a Markov chain Monte Carlo algorithm using the model-based geostatistics framework of Diggle et al. (1998). For the particular application considered in this paper the specification of prior distributions for the model parameters was done along the lines of the work described by Alexander et al. (2000) and Diggle et al. (2002). Specifically, the prior distribution for  $k$  was chosen to be gamma with density proportional to  $k^{(1.5-1)} \exp(-0.01k)$  so that, *a priori*,  $k$  had a high variance. We chose independent improper uniform prior distributions for all the regression coefficients associated with the covariates and the constant term. For the parameters  $\sigma^2$  and  $\alpha$  we adopted the following vague priors:  $f(\sigma^2) \propto 1/\sigma^2$  and  $f(\alpha) \propto 1/\alpha^2$ . In the absence of any prior knowledge about the model parameters, the choice of non-informative improper priors was dictated by practical considerations, e.g. convergence issues and allowing the data to have a greater influence in determining the posterior.

We simulated realizations from the posterior distributions by means of a single-component Metropolis-Hastings algorithm. The parameters were updated by using a random-walk Metropolis algorithm with a Gaussian proposal density for the regression parameters,  $\log(\sigma^2)$ ,  $\log(\alpha)$  and  $\log(k)$ . At each iteration, the updating of  $\sigma^2$  and  $\alpha$  required the inversion of the  $n \times n$  covariance matrix of the spatial effect, where  $n$  is the number of locations. In our example,  $n = 459$ . The Markov chain was run for 110,000 iterations. After an initial burn-in of 10,000 iterations, the chain was sampled every 20<sup>th</sup> iteration to yield a sample of 5000 values from the posterior distribution of each of the model parameters. The convergence was judged by looking at the MCMC traces of the model parameters and also by computing the Monte Carlo errors. The traces differed in the extent to which they showed good mixing, but they all exhibited a reasonable degree of convergence to a stationary distribution. Spatial prediction was undertaken on a fine grid of 3304 locations with a grid-spacing of 12 km. At each prediction location 5000 values were simulated using the method outlined in Diggle et al. (1998). The Bayesian inference and prediction were undertaken using a program custom-written in the C language.

## RESULTS

Intensity data were available for 31,458 school children (15,904 boys and 15,554 girls) aged 5-22 years (mean age 11.5 years, median age 12 years, 90% aged between 6-16 years). The median prevalence of *S. mansoni* infection was 3.3% ranging from 0-97.2% by school. The overall mean egg count was 116 epg (ranging 0.9 to 3234 eggs/gram faeces by school) (Table 1). Figure 1 displays the spatial variation in infection intensity and shows that intensity was highest along the shores of the major lakes and in Uganda, along the river Nile.

The results of the Bayesian geostatistical model are presented in Table 2. Model coefficients indicate that females had significantly lower egg counts than males and that higher egg counts are found in areas at low elevations and close to perennial water. The posterior mean of  $0.137 * 122 \approx 16.7$  km for  $\alpha$ , the range of spatial correlation, corresponds to a 'half-distance' for correlation of  $16.7 * \log 2 \approx 11.6$  km, with  $3 * 16.7 \approx 50.1$  km being the distance at which the correlation reduces to 0.05. The 95% posterior interval for  $\alpha$  of (11.1, 24.2) km indicates that the residual spatial variation operates on a small scale. This is in agreement with the empirical variogram of the mean number of eggs in each location in Figure 2a which exhibits a rising trend up to a distance of around 40 km. The posterior mean value of the overdispersion parameter was 0.060, confirming that the data were highly overdispersed even after accounting for spatial correlation. In addition, the estimate of the overdispersion parameter for the non-spatial model was 0.031, suggesting that inclusion of the spatial random-effect accounted for some of the overdispersion and, despite similar posterior mean values, the Bayesian credible intervals were narrower for the non-spatial model than the spatial model. All the parameters have mostly symmetric posterior distributions excepting  $\sigma^2$  and  $\alpha$  which are slightly positively skewed. The empirical variogram of the Pearson residuals from the fitted non-spatial regression model in Figure 2b is quite similar to the variogram in Figure 2a. This indicates that the non-spatial regression model is inadequate in explaining the short-range spatial structure, whereas the variogram of the residuals from the spatial model shown in Figure 2c is essentially flat.

The map of the posterior mean predicted intensity value is presented in Figure 3. Model predictions of infection intensity indicate a high intensity in areas adjacent to Lakes Victoria, Kyoga, Albert and the northern part of Lake Edward as well as along the Nile River valley in Uganda. Low intensity areas were predicted in northeast and southwest Uganda and much of northwest Tanzania away from the lakeshore. Figure 4 presents the standard deviations of the modelled intensities. As expected, a lower level of uncertainty was apparent in locations close to observed data points and a higher level of uncertainty was apparent in locations distant from observed data points. Since observed data were well-distributed across the region, most areas

were characterised by a reasonable level of prediction certainty. An exception to this was northeast Uganda and areas between Lakes Victoria, Albert and Edward in western Uganda, areas which are too arid and too high, respectively to support transmission of *S. mansoni*, and therefore were not sampled.

## DISCUSSION

This study presents a novel application of Bayesian geostatistical modelling to predict intensity of infection with *S. mansoni*, a highly aggregated epidemiologic index. Results identify the role of environmental risk factors in explaining geographical heterogeneity in infection intensity and show how these factors can be used to develop a predictive map. Such a map provides an empirical basis for identifying priority areas when implementing control and for predicting the potential impact of control.

The observed risk factors are already well known and are consistent and interpretable with the epidemiology of infection and known biology of freshwater snails, the intermediate hosts for *S. mansoni*. Malacological studies confirm *Biomphalaria* snail species in East Africa tend to prefer large water body habitats (Prentice et al., 1972; Lwambo et al., 1999). Furthermore, individuals living on or near the shore are more likely to undertake risky water contact behaviour, increasing their exposure to infection. Observed differences among males and females may also be exposure-related, although it is also suggested that hormonal differences between males and females may account for higher infection rates in males than females (Webster et al., 1997). The role of other host-specific factors, such as genetics, immunity, nutrition, hygienic behaviour and education, as risk factors is also recognised. Climatic conditions, primarily temperature and rainfall, have been shown to influence the distribution and density of snails and the rate of schistosomal development in the snail host (Appleton, 1978; Sturrock, 1993). At high altitudes, low temperatures cause snails to die before the cercariae mature from sporocysts, limiting transmission (Pflüger, 1980).

Our predictive maps of schistosome transmission are less affected by seasonality than other, more seasonally dynamic, parasitic diseases, such as malaria. Although seasonal dynamics in schistosome transmission may occur, such fluctuations may be of little significance to the overall parasite equilibrium within communities. This is because the lifespan of adult worms is typically much longer (1-10 years) than the periods in the year during which the basic reproductive number ( $R_0$ ) is less than unity, and  $R_0$  will on average be greater than one, maintaining overall endemicity (Anderson, 1982).

Previous studies predicting parasitic diseases in Africa on the basis of published data suffer from certain methodological problems including dissimilar age groupings of surveys and use of different diagnostic techniques to assess infection (e.g. Kleinschmidt et al. 2001). The inclusion of survey data from comparable age groups collected using the same diagnostic method (Kato-Katz quantitative smear) reduces this inherent variability. However, methodological problems related to the limited sensitivity and specificity of single stool samples to detect light infection (de Vlas et al., 1993) constitutes a limitation of the current analysis and may widen the prediction errors by introducing greater variability into the estimates of infection intensity.

Our approach highlights a number of key methodological limitations and areas for further work. First, the current model does not account for anisotropic and non-stationary spatial variation. Data is currently being collected in other countries with SCI-supported control programmes, including Zambia in southern Africa and Burkina Faso, Mali and Niger in west Africa. These data cover wide areas with marked environmental diversity. Hence, there is scope of investigating the possible presence of anisotropy and non-stationarity in observed spatial variation. Second, there is the possibility that the degree of parasite aggregation may vary

between endemic populations, reflecting area-specific differences in host exposure and immunity (Guyatt et al., 1994). We did not observe any variation in aggregation patterns with host age, presumably reflective of the relatively narrow age range of individuals included in the analysis, or according to environmental covariates. Third, zero-truncated negative binomial distribution has previously been used to describe patterns of *Wuchereria bancrofti* (Grenfell et al., 1990), *Onchocerca volvulus* (Filipe et al., 2005) and *Loa loa* (Pion et al., 2006) and may provide an alternative approach to modelling aggregated distributions. This would involve using a mixture model, i.e. a negative binomial distribution with an extra point mass at zero. Over the next few years improved approaches to modelling geostatistical processes and transmission dynamics will iteratively improve our ability to predictive disease patterns for geographical targeting of interventions.

As well as identifying areas where population-based morbidity control, using praziquantel, is most warranted (Brooker and Michael, 2000; Brooker, 2002), spatial predictions of infection intensity can also help define the frequency of treatment required to reduce morbidity and to model the potential impact of treatment. Mathematical models of transmission dynamics (Chan et al., 1995) use mean intensity of infection to predict rates of re-infection over fixed time intervals, and provide estimates of infections and heavy infections prevented each year and the number of morbidity cases/year prevented. The integration of Bayesian geostatistical modelling with transmission dynamics models will provide an important analytical platform to help better understand the epidemiology of schistosomiasis and provide a more robust evidence-base approach to implementing and evaluating control.

#### ACKNOWLEDGEMENTS

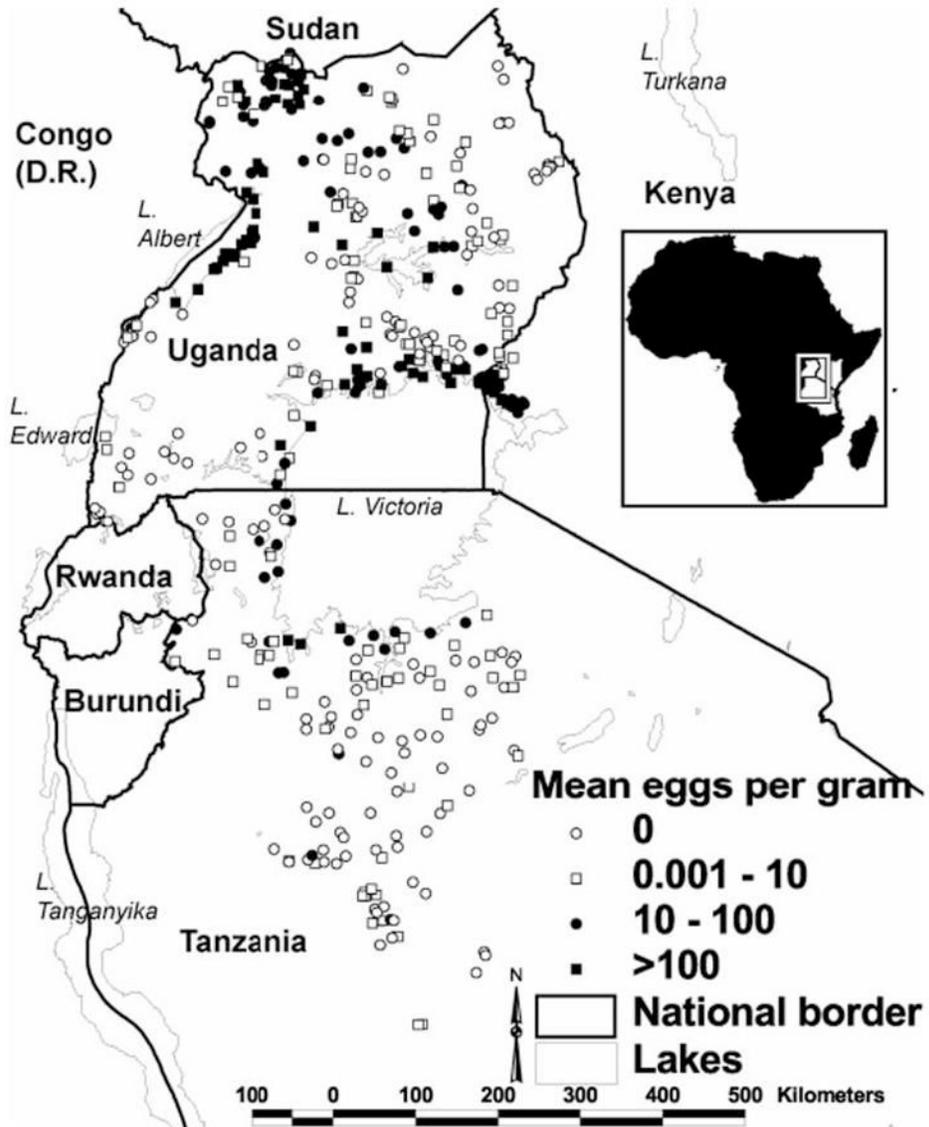
This project was supported by the Schistosomiasis Control Initiative which is generously supported by the Bill and Melinda Gates Foundation. Simon Brooker is supported by a Wellcome Trust Advanced Training Fellowship (073656). We thank the many field workers who carried out the field surveys. We also thank Drs Narcis Kabateriene, Nicholas Lwambo, Sian Clarke, Benson Estambale and Njagi Kiamba for providing access to the survey data, and Professor Alan Fenwick for his support and contributions.

#### REFERENCES

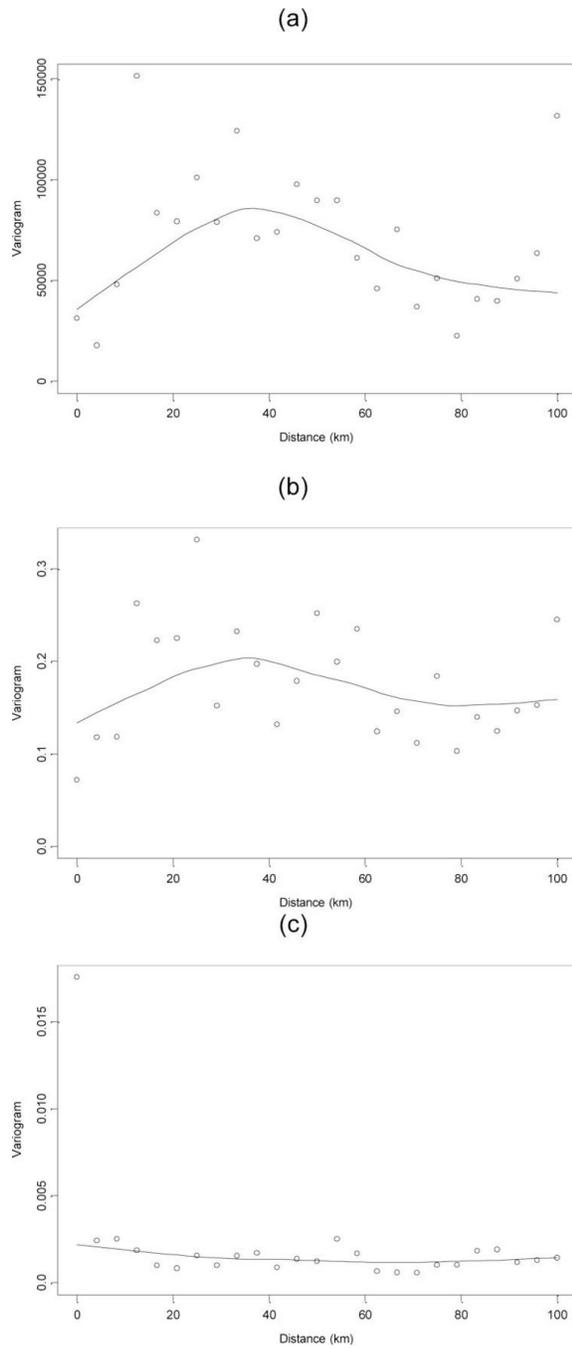
- Alexander N, Moyeed R, Stander J. Spatial modelling of individual-level parasite counts using the negative binomial distribution. *Biostatistics* 2000;1:453–463. [PubMed: 12933567]
- Alexander ND, Moyeed RA, Hyun PJ, Dimber ZB, Bockarie MJ, Stander J, Grenfell BT, Kazura JW, Alpers MP. Spatial variation of Anopheles-transmitted *Wuchereria bancrofti* and *Plasmodium falciparum* infection densities in Papua New Guinea. *Filaria Journal* 2003;2:14. [PubMed: 14525619]
- Anderson, RM. Epidemiology. In: Cox, FEG., editor. *Modern Parasitology*. Blackwell; Oxford: 1993. p. 75-166.
- Anderson, RM.; May, RM. *Infectious Diseases of Humans: dynamics and control*. Oxford University Press; Oxford, UK: 1991.
- Anderson, RM. The population dynamics and control of hookworm and roundworm infection. In: Anderson, RM.; Anderson, RM., editors. *Population Dynamics of Infectious Diseases: Theory and Applications*. Chapman and Hall; London: 1982. p. 67-109.
- Appleton CC. Review of Literature on abiotic factors influencing the distribution and life-cycles of Bilharziasis intermediate host snails. *Malacological Review* 1978;11:1–25.
- Basáñez M-G, Boussinesq M. Population biology of human onchocerciasis. *Philosophical Transactions of the Royal Society B: Biological Sciences* 1999;354:809–826.
- Brooker S, Michael E. The potential of geographical information systems and remote sensing in the epidemiology and control of human helminth infections. *Advances in Parasitology* 2000;47:245–288. [PubMed: 10997209]
- Brooker S. Schistosomes, snails and satellites. *Acta Tropica* 2002;82:209–216.

- Brooker S, Miguel EA, Moulin S, Waswa P, Namunyu R, Guyatt H, Bundy DAP. The potential of rapid screening methods for *Schistosoma mansoni* in Western Kenya. *Annals of Tropical Medicine and Parasitology* 2001;95:343–351. [PubMed: 11454244]
- Brooker S, Clements ACA, Bundy DAP. Global epidemiology, ecology and control of soil-transmitted helminth infections. *Advances in Parasitology* 2006;62:223–265.
- Carabin H, Escalona M, Marshall C, Vivas-Martinez S, Botto C, Joseph L, Basáñez M-G. Prediction of community prevalence of human onchocerciasis in the Amazonian onchocerciasis focus: Bayesian approach. *Bulletin of World Health Organization* 2003;81:482–490.
- Clements A, Lwambo N, Blair L, Nyandindi U, Kaatano G, Kinunghi S, Webster J, Fenwick A, Brooker S. Bayesian spatial analysis and disease mapping: tools to enhance planning and implementation of a schistosomiasis control programme in Tanzania. *Tropical Medicine and International Health* 2006;11:490–503. [PubMed: 16553932]
- Chan MS, Guyatt HL, Bundy DA, Booth M, Fulford AJ, Medley GF. The development of an age structured model for schistosomiasis transmission dynamics and control and its validation for *Schistosoma mansoni*. *Epidemiology and Infection* 1995;115:325–344. [PubMed: 7589272]
- Clarke, S.; Njagi, K.; Jukes, MCH.; Estambale, B.; Khasakhala, L.; Ajanga, A.; Otido, J.; Ochola, S.; Magnussen, P.; Brooker, S. Intermittent preventive treatment in schools: malaria parasitaemia, anaemia and school performance [MIM-KN-409248]. Meeting abstract. *Acta Tropica*; 4th MIM Pan-African Malaria Conference; 2005. p. S133-S134.
- de Vlas SJ, Nagelkerke NJ, Habbema JD, van Oortmarssen GJ. Statistical models for estimating prevalence and incidence of parasitic diseases. *Statistical Methods in Medical Research* 1993;2:3–21. [PubMed: 8261249]
- Diggle P, Tawn JA, Moyeed R. Model-based geostatistics. *Applied Statistics* 1998;47:299–350.
- Diggle P, Moyeed R, Rowlingson B, Thompson M. Childhood malaria in the Gambia: a case-study in model-based geostatistics. *Applied Statistics* 2002;51:493–506.
- Filipe JAN, Boussinesq M, Rez A, Collins AC, Vivas-Martinez S, Grillet M-G, Little MP, Basáñez M-G. Human infection patterns and heterogeneous exposure in river blindness. *Proceedings of the National Academy of Science, USA* 2005;102:15265–15270.
- Gemperli A, Vounatsou P, Kleinschmidt I, Bagayoko M, Lengeler C, Smith T. Spatial patterns of infant mortality in Mali: the effect of malaria endemicity. *American Journal of Epidemiology* 2004;159:64–72. [PubMed: 14693661]
- Gemperli A, Vounatsou P, Sogoba N, Smith T. Malaria mapping using transmission models: application to survey data from Mali. *American Journal of Epidemiology* 2006;163:289–297. [PubMed: 16357113]
- Grenfell BT, Das PK, Rajagopalan, Bundy DAP. Frequency distribution of lymphatic filariasis microfilariae in human population: population processes and statistical estimation. *Parasitology* 1990;101:417–427. [PubMed: 2092297]
- Guyatt HL, Smith T, Gryseels B, Lengeler C, Mshinda H, Siziya S, Salanave B, Mohome N, Makwala J, Ngimbi KP, Tanner M. Aggregation in schistosomiasis: comparison of the relationships between prevalence and intensity in different endemic areas. *Parasitology* 1994;109:45–55. [PubMed: 8058368]
- Hay, SI.; Randolph, SE.; Rogers, DJ. *Advances in Parasitology*. 47. Academic Press; London: 2000. Remote sensing and geographic information systems in epidemiology.
- Hay, SI.; Graham, AJ.; Rogers, DJ. *Advances in Parasitology*. 62. Academic Press; London: 2006. Global mapping of infectious diseases: methods, examples and emerging applications.
- Kabatereine NB, Brooker S, Tukahebwa EM, Kazibwe F, Onapa A. Epidemiology and geography of *Schistosoma mansoni* in Uganda: implications for planning control. *Tropical Medicine and International Health* 2004;9:372–380. [PubMed: 14996367]
- Kleinschmidt I, Omumbo J, Briet O, van de Giesen N, Sogoba N, Mensah NK, Windmeijer P, Moussa M, Teuscher T. An empirical malaria distribution map for West Africa. *Tropical Medicine and International Health* 2001;6:779–786. [PubMed: 11679126]
- Lwambo NJS, Siza JE, Brooker S, Bundy DAP, Guyatt H. Patterns of concurrent infection with hookworm and schistosomiasis in school children in Tanzania. *Transactions of the Royal Society and Tropical Medicine* 1999;93:497–502.

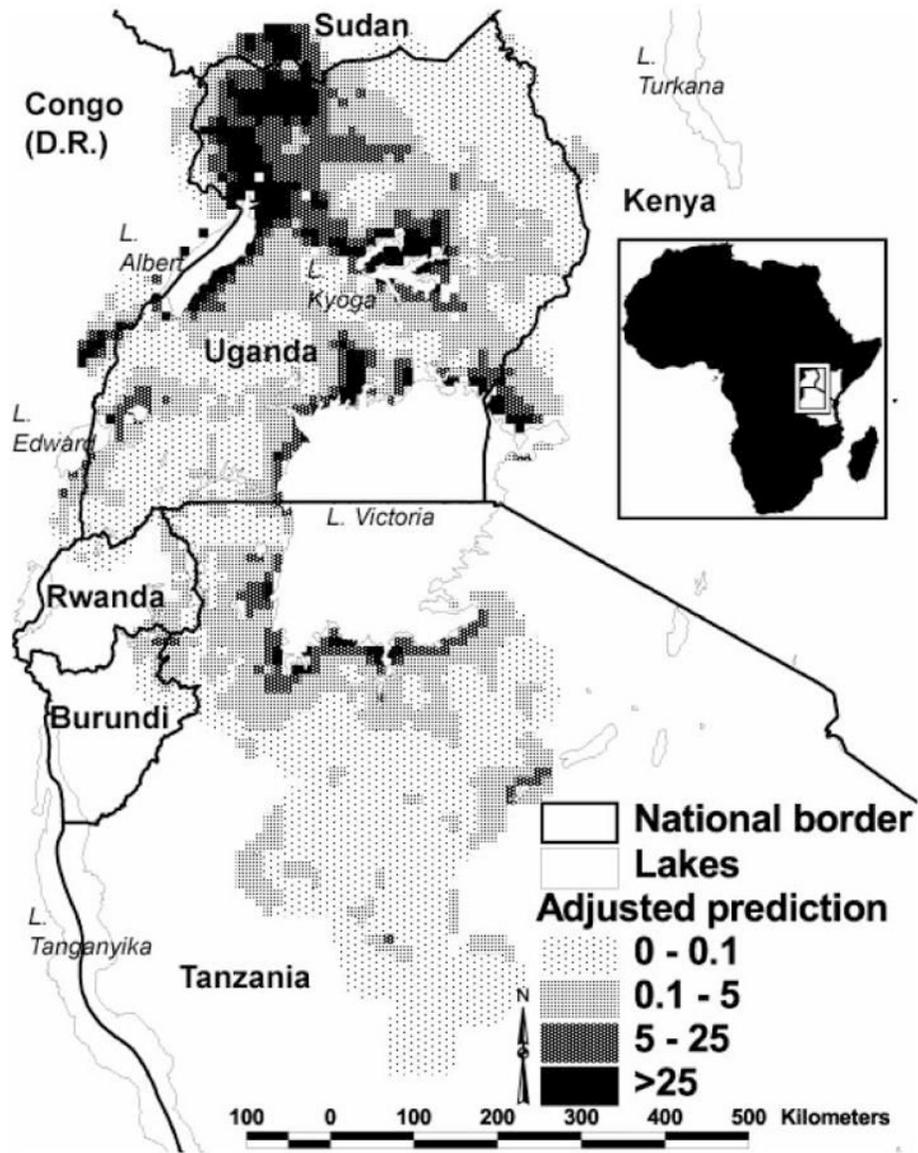
- Nodtvedt A, Dohoo I, Sanchez J, Conboy G, DesCjteaoux L, Keefe G, Leslie K, Campbell J. The use of negative binomial modelling in a longitudinal study of gastrointestinal parasite burdens in Canadian dairy cows. *Canadian Journal of Veterinary Research* 2002;66:249–257. [PubMed: 12418780]
- Pflüger W. Experimental epidemiology of schistosomiasis I. The prepatent period and cercarial production of *Schistosoma mansoni* in *Biomphalaria* snails at various constant temperatures. *Zeitschrift für Parasitenkunde* 1980;63:159–169.
- Pion SDS, Filipe JAN, Kamgno J, Gardon J, Baséñez M-G, Boussinesq M. Microfilarial distribution of *Loa loa* in the human host: population dynamics and epidemiological implications. *Parasitology* 2006;131:1–12.
- Prentice MA. Distribution, prevalence and transmission of schistosomiasis in Uganda. *Uganda Medical Journal* 1972;1:136–139.
- Raso G, Matthys B, N'Goran EK, Tanner M, Vounatsou P, Utzinger J. Spatial risk prediction and mapping of *Schistosoma mansoni* infections among schoolchildren living in western Côte d'Ivoire. *Parasitology* 2005;131:97–108. [PubMed: 16038401]
- Raso G, Vounatsou P, Singer BH, N'Goran EK, Tanner M, Utzinger J. An integrated approach for risk profiling and spatial prediction of *Schistosoma mansoni*-hookworm coinfection. *Proceedings of the National Academy of Science U S A* 2006;103:6934–6939.
- Shaw DJ, Grenfell BT, Dobson AP. Patterns of macroparasite aggregation in wildlife host populations. *Parasitology* 1998;117:597–610. [PubMed: 9881385]
- Sturrock, RF. The Intermediate Hosts and Host- Parasite Relationships. In: Jordan, P.; Webbe, G.; Sturrock, RF., editors. *Human schistosomiasis*. CAB International; Wallingford: 1993. p. 33-85.
- Tatem AJ, Noor AM, Hay SI. Assessing the accuracy of satellite derived global and national urban maps in Kenya. *Remote Sensing of Environment* 2005;96:87–97.
- Thogmartin WE, Sauer JR, Knutson MG. A hierarchical spatial model of avian abundance with application to cerulean warblers. *Ecological Applications* 2004;14:1766–1779.
- Thomson MC, Connor SJ, D'Alessandro U, Rowlingson B, Diggle P, Cresswell M, Greenwood B. Predicting malaria infections in Gambian children from satellite data and bed net use surveys: the importance of spatial correlation in the interpretation of results. *American Journal of Tropical Medicine and Hygiene* 1999;61:2–8. [PubMed: 10432046]
- Webster M, Correa-Oliveira R, Gazzinelli G, Viana IR, Fraga LA, Silveira AM, Dunne DW. Factors affecting high and low human IgE responses to schistosome worm antigens in an area of Brazil endemic for *Schistosoma mansoni* and hookworm. *American Journal of Tropical Medicine and Hygiene* 1997;57:487–494. [PubMed: 9347969]
- Woolhouse MEJ, Etard J-F, Dietz K, Ndhlovu PD, Chandiwana SK. Heterogeneities in schistosome transmission dynamics and control. *Parasitology* 1998;117:475–482. [PubMed: 9836312]
- World Health Organization. Report of the WHO informal consultation on schistosomiasis control. WHO/CDS/CPC/SIP/99.2. WHO; Geneva: 1999.
- Yang GJ, Vounatsou P, Zhou XN, Tanner M, Utzinger J. A Bayesian-based approach for spatio-temporal modeling of county level prevalence of *Schistosoma japonicum* infection in Jiangsu province, China. *International Journal of Parasitology* 2005;35:155–162. [PubMed: 15710436]
- Yapi YG, Briet OJ, Diabate S, Vounatsou P, Akodo E, Tanner M, Teuscher T. Rice irrigation and schistosomiasis in savannah and forest areas of Côte d'Ivoire. *Acta Tropica* 2005;93:201–211. [PubMed: 15652334]
- Zheng T, Salganik MJ, Gelman A. How many people do you know in prison?: Using overdispersion in count data to estimate social structure in networks. *Journal of American Statistical Association* 2006;101:409–423.



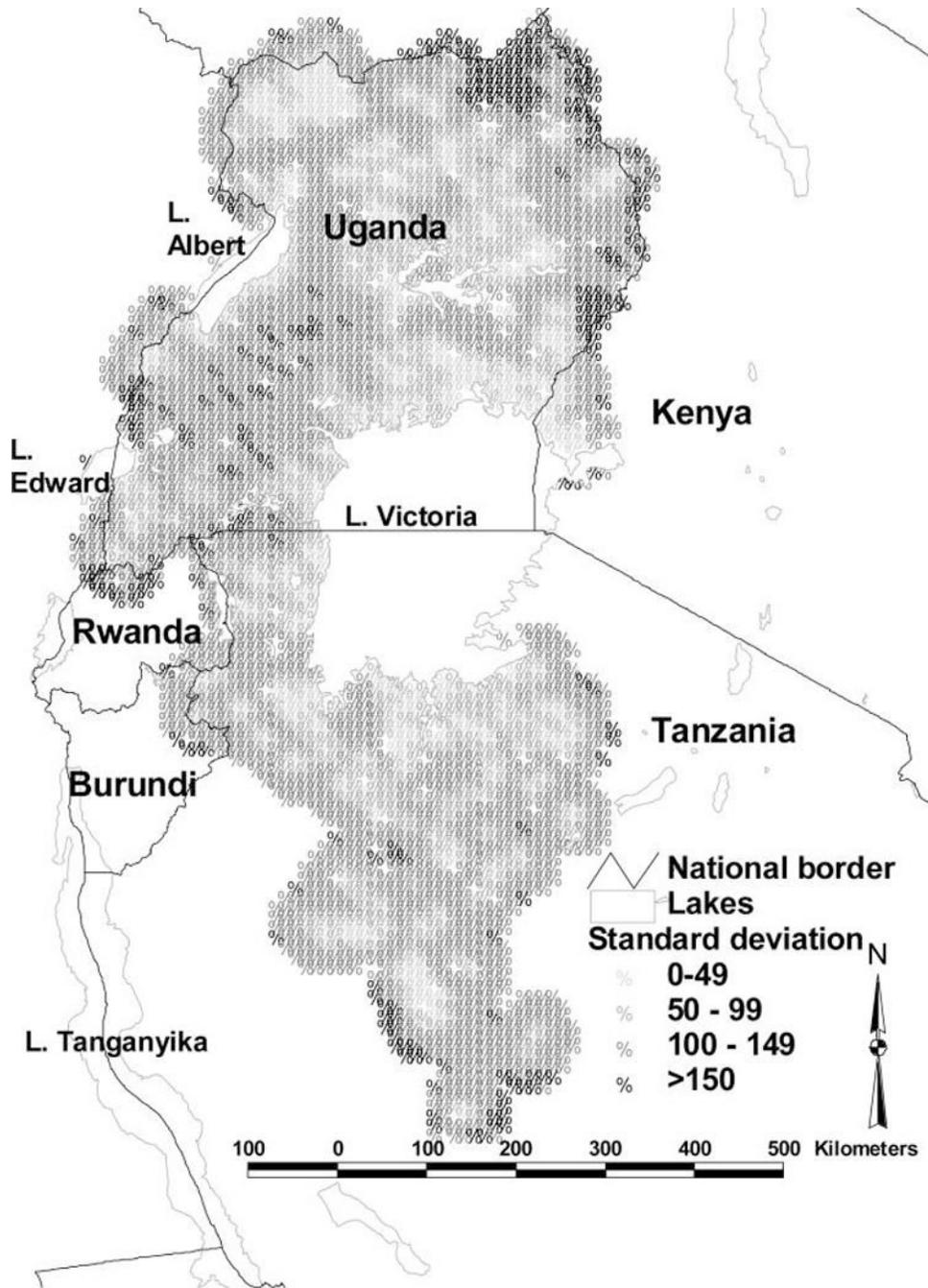
**Figure 1.** Distribution of school surveys and observed intensity of *Schistosoma mansoni* (as measured by the mean number of eggs per gram of faeces in each location) in 31,458 school children aged 5-22 years at 459 locations in East Africa conducted between 1999 and 2004. Locations of major lakes are also indicated.



**Figure 2.** (a) Empirical variogram of the mean number of eggs in each location. (b) Empirical variogram of the average of the Pearson residuals for each location from the non-spatial model. (c) Empirical variogram of the average of the Pearson residuals for each location from the spatial model. The curves were produced using a loess smoother.



**Figure 3.** Predicted intensity of infection (eggs/gram faeces) with *Schistosoma mansoni* in East Africa, adjusted for covariates (distance to perennial water body and elevation).



**Figure 4.** Posterior standard deviations of the model predictions of the intensity of infection with *Schistosoma mansoni* in East Africa.

**Table 1**

Summary of data on *Schistosoma mansoni* in East Africa used in the analysis.

Country	Number of schools	Number of children	Mean age, years (range)	% male	Overall prevalence (range by school)	Mean intensity (range by school)	Reference
Kenya	21	1,091	12.9 (10-20)	52.8	14.4 (0-30)	51.4 (0-141)	Clarke et al. (2005)
Kenya	24	1,6775	13.3 (6-20)	53.9	32.3 (0.2-97)	220.4 (3-1774)	Brooker et al. (2001)
Tanzania	143	8,617	14.0 (6-22)	49.6	4.4 (0-95)	17.6 (0-1384)	Clements et al. (2006)
Uganda	271	20,073	10.4 (5-21)	50.1	16.4 (0-97)	154.2 (0-3234)	Kabaterere et al. (2004)
Total	459	31,458	11.5 (5-22)	50.6	18.6	116.7 (0.9-3234)	

**Table 2**

Coefficients of the Bayesian negative binomial geostatistical model based on overdispersed individual *Schistosoma mansoni* egg count data, obtained from 31,458 school-age children and young adults in East Africa during 2003-2004.

Parameter	Spatial Model				Non-spatial Model			
	Posterior Mean	95% Bayesian CI			Posterior Mean	95% Bayesian CI		
		Lower 2.5%	Upper 97.5%	Posterior Median		Lower 2.5%	Upper 97.5%	Posterior Median
Intercept	9.941	5.768	13.223	10.055	10.295	9.826	10.770	10.292
Sex (Female) <sup>*</sup>	-0.413	-0.721	-0.106	-0.414	-0.463	-0.590	-0.336	-0.463
Elevation	-0.007	-0.010	-0.004	-0.007	-0.005	-0.005	-0.004	-0.005
Overdispersion parameter	0.060	0.058	0.062	0.060	0.031	0.030	0.032	0.031
Distance to nearest perennial water body <sup>†</sup>	-5.358	-7.512	-3.295	-5.364	-4.187	-4.334	-4.041	-4.188
Variance of the spatial random-effect	24.362	19.062	32.070	23.955				
Range of spatial correlation <sup>‡</sup>	0.137	0.091	0.198	0.134				

<sup>\*</sup> Reference category: Sex (Male).

<sup>†</sup> Unit: decimal degrees. 1 decimal degree = approximately 122 km at the equator.